

ILOs (intended learning outcomes): Student should be able to use microscope for examination of bacterial stained smears.

- **Study the different parts of the microscope.**
 - The support system of the microscope (body, base and stage).
 - The illumination system (mirror, condenser and iris diaphragm).
 - The magnifying system (eye piece lens, objective lenses).
 - The adjusting system (coarse and fine adjustment knobs).
- **Magnification:**
 - The magnification achieved by a microscope is a product of the magnifying power of the eye piece and the objective lenses (low power, high power and oil immersion lenses).
 - Calculate the magnification of the microscope when using:
 - Low power lens: $10 \times 40 = 400$ times.
 - High power lens: $10 \times 100 = 1000$ times.
 - Oil-immersion lens: $10 \times 100 = 1000$ times.
- **Resolution:**
 - The resolving power of the microscope is its capacity to distinguish two neighboring points as separate entities.
 - It depends on:
 1. Wave length of light
 2. Numerical aperture of objective length
 - What is the smallest size that ordinary microscope can visualize?
200nm
 - How to examine a stained film by oil-immersion lens?
 1. Put the slide on microscope
 2. use oil over slide
 3. use the coarse then fine
 4. look at eye piece

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Report(2): Preparing a smear for simple staining?

- ILOs: 1. To prepare a smear for staining.
2. To perform simple stain.

- How to prepare a smear for staining?

- sterilized bacteriology loop by heat
- wait for the loop to be cold quickly
- take a drop of water and put it on slide
- sterilize the loop again
- take a drop of specimen by loop and in the slide and spread
- dry the slide and fix it by flame

- List steps of simple staining by methylene blue?

- Put methylene blue after preparation
- wait for minute then wash it by water, dry slide then put
- a drop of oil then examine by immersion lens

- Examine the stained smear under oil-immersion lens (OIL) of the microscope.

✓ setup the microscope against a good source for light
place one drop of immersion oil on the slide
put and examine it by immersion lens

- Description of the stained bacteria.

- Morphology: bacilli

- Arrangement: clusters?

- Drawing of the bacteria:



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Port (3): Gram staining

- ILOs:**
1. To perform Gram stain procedure.
 2. To examine and identify bacteria of different morphology and Gram reaction.

- List the steps of Gram staining:

- Cover the smear with crystal violet or methy violet for (30-60) Sec.
 - Pour it off and wash with water
 - add Iodine solution and leave it to act for 1 min then pour it off and wash with water
 - Decolorize by adding 95% alcohol or colour alcohol
 - reply till no violet colour comes off the smear
 - Wash rapidly with water
 - Counter stain with safranin or dilute basic fuchsin for 1 min
 - Wash with water then place the slide to air dry
- How do Gm +ve and Gm -ve bacteria behave during the following staining steps?

| | Gm +ve | Gm -ve |
|--|-----------------------|-------------|
| Staining with methyl violet | ✓ | ✓ |
| Washing with alcohol | resist decolorization | decolorized |
| Staining with the counter stain (diluted carbol fuchsin) | violet | red |

- Examine the Gm stained smear under OIL of the microscope.
- Setup the microscope facing a good source light
Place small drop of immersion oil on slide
Put the slide on the stage of microscope and use oil immersion lens and focus on object using fine adjustment
- Description of the Gm stained bacteria:
- Morphology: Bacilli
 - Arrangement: clusters no special arrangement
 - Gm reaction: -ve
 - Drawing of the bacteria:

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Report (4): Sterilization and disinfection

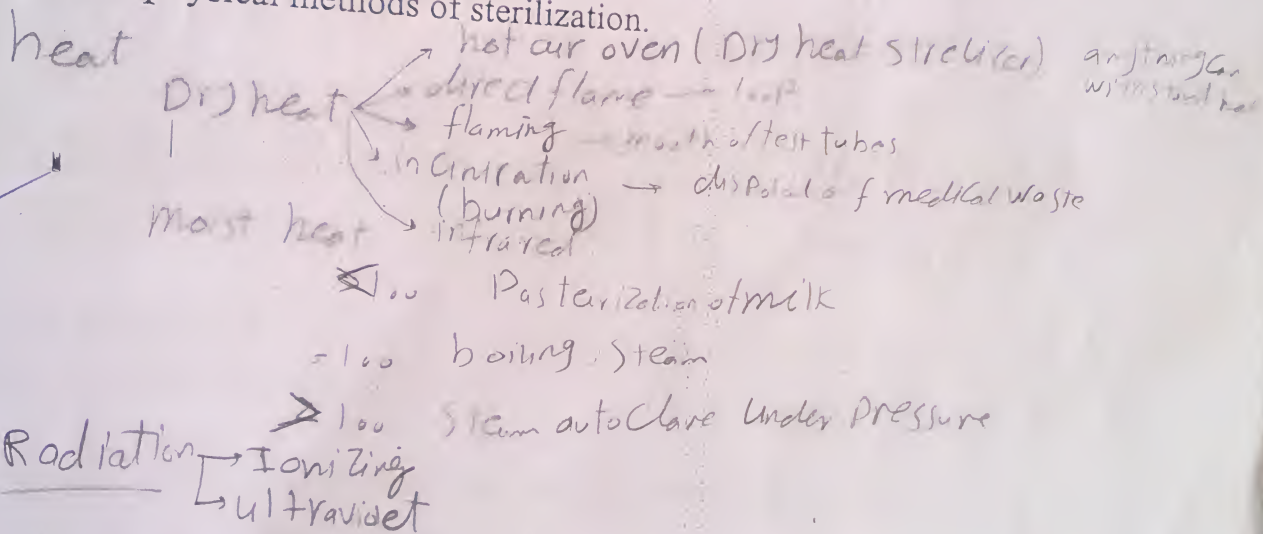
ILOs: To examine some tools of physical method of sterilization (simple autoclave, hot air oven and seitz filter)

- Sterilization is: Killing of all signs of life including spores
 Pathogenic, non Pathogenic

• Methods:

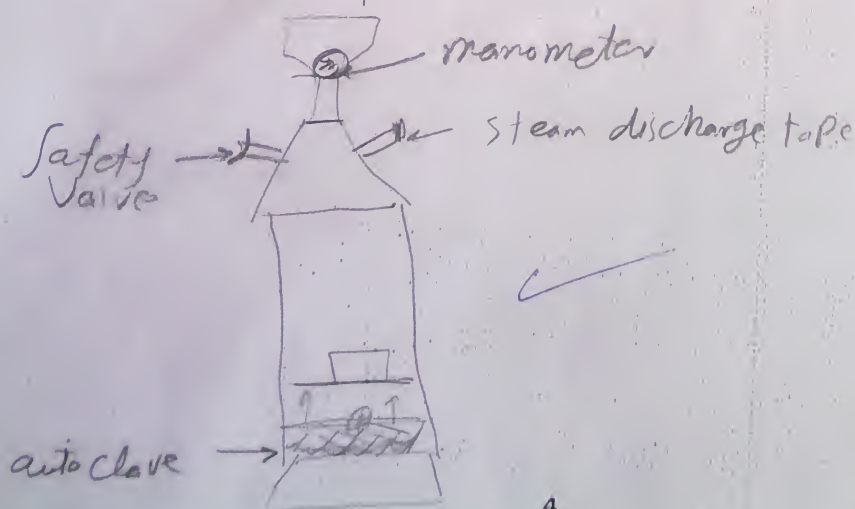
1. Physical \rightarrow heat \rightarrow radiation
2. Chemical \rightarrow disinfectant, antiseptic
3. Gaseous \rightarrow ethylene oxide
4. Filtration \rightarrow gas plasma

• Outline physical methods of sterilization.



Filtration

- Draw a labeled diagram of simple autoclave.



What is the principle of plasma-gas sterilizer?
 H_2O_2 alone or with Peracetic acid excited by radio freq. energy
 → free radicals toxic to bacteria

- Filtration is the suitable sterilization method for: sterilization
 biological fluids (serum, blood, Plasma)

Explain why? destroyed by heat

- Mention types of filters:
 - multiPore filter (Pore size 15 μm)
 - Syringe filter (small volumes)
 - Vacuum filter
- Seitz filter
 air filter (HEPA)
- Ionizing radiation method is mainly used for
 Plastic devices (gloves, Catheters, Sutures)

- Disinfection is killing of most pathogenic organisms
not including endospores

- Mention examples of chemical disinfectants and their uses:

Protein denaturation

alcohol (ethyl alcohol) 70% → Protein denaturation

Phenols (QAC)

inactivation of enzymes

Chlorine $\left\{ \begin{array}{l} \text{gas} \rightarrow \text{water} \\ \text{liquid} \rightarrow \text{hospital/home} \end{array} \right.$

oxidizing

(Mordant Sterilizer)
 • Formaldehyde
 • Glutaraldehyde

Detergent $\left\{ \begin{array}{l} \text{anionic} \rightarrow \text{Soap} \\ \text{Cationic} \rightarrow \text{QAC} \end{array} \right.$

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Praxis

→ electron microscope

Report (5): Demonstration of different culture media.

ILOs: To examine and identify different culture media.

1. Simple media

- Complete the following table (1)

| Medium | Main component | Use |
|---------------|------------------------------|--|
| Peptone water | Water - Peptone - NaCl | - Production of other media - test for indole |
| N. broth | Peptone water + meat extract | - Coagulase test - blood Culture |
| N. agar | Peptone water + Agar | - Isolation of organism - other media |

2. Enriched media: (contain highly nutritive substances as blood, serum, egg)

- Complete the following table (2)

| Medium | How to prepare & sterilize | Use |
|-----------------|---|--|
| Blood agar | • Steam autoclave under pressure at 121°C for 20 hrs (nutrient agar) • Blood at 55°C (Blood st. by filtration) | - Isolation of bacteria (fastidious) - as Indicator media (type of hemolysis) |
| Chocolate agar | as above but blood added at 100°C (hematin) | growth factor for <i>Neisseria</i> , <i>Haemophilus</i> (fastidious) |
| Löffler's serum | 1/75 Serum + 1/25 glucose | Isolation of diphtheria |

3. Selective media: (contain selective agent(s) that inhibits all but not the selected bacteria).
- Complete the following table (3)

| Medium | Selective agent(s) | Grown bacteria |
|-------------------|-----------------------|----------------|
| Lowenstein Jensen | malachite green | T.B. |
| TCBS | Potassium tellurite | diphtheria |
| MSA | modified chelate agar | Gonorrhoea |

4. Differential (Indicator media)

- Complete the following table (4)

| Medium | Test sugar | Indicator | Different colours of growing colonies |
|-----------|------------|----------------|---------------------------------------|
| MacConkey | Lactose | neutral red | - yellow - Pink |
| TCBS | Sucrose | promethol blue | - Yellow - Green |
| MSA | Mannitol | Phenol red | - Yellow - Pink |

Media for anaerobic bacteria:

- Robertson's Cooked meat
- Thioglycollate broth

- Anaerobic Gas Pack system:

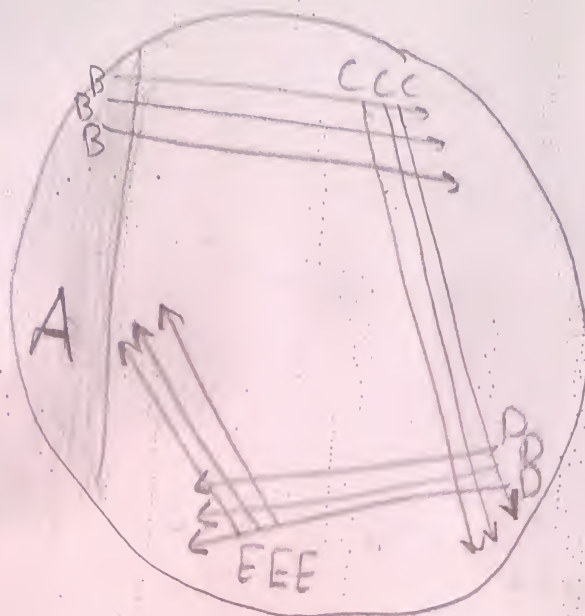
How the anaerobic atmosphere (or microaerophilic) is generated by gas pack system inside the anaerobic jar?

The hydrogen is generated inside the jar by placing a special Gas Pack envelope immediately before placing it in the jar, it will release hydrogen & CO₂. The presence of the Cold Catalyst in the jar allows the hydrogen released to combine with the oxygen in the jar to give strictly anaerobic condition.

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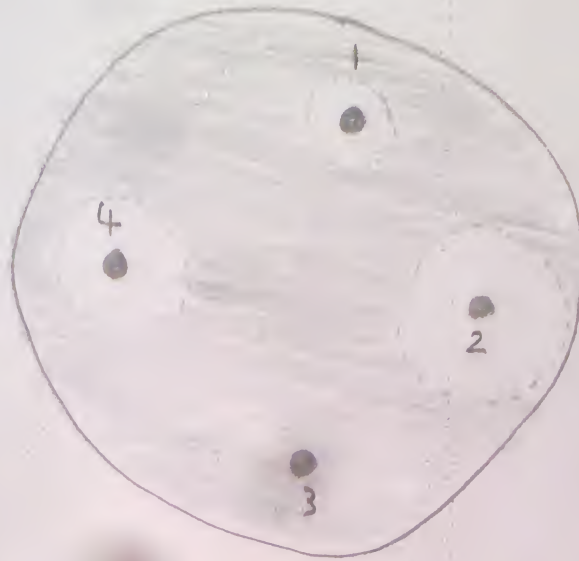
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- technique for isolation of bacteria.
- Interpret antibiotic sensitivity test.
- Draw a diagrammatic description of plating out technique.



- How to get a pure culture?
 - 1st Culture
 - 2nd Culture
 - Tertiary
 - S Shaped
 - Colony
- Pure culture is used for:
 - Identification morphology character
 - antibiotic sensitivity test
 -

gram describing it. The sensitivity plate *showing...* and draw a *Zone of inhibition*



- The plate showing a zone of inhibition around the discs No. 2 ~~4~~ and 4. The discs No. 3 show no inhibition zone.
- Complete the following table:

| The antibiotic disc No. | Susceptibility of bacteria |
|-------------------------|----------------------------|
| 1 | Low |
| 2 | High |
| 3 | No |
| 4 | Medium |
| | |
| | |

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Demonstration of bacterial culture characteristics

ILOs: To examine and identify different selected culture characteristics of bacteria.

- Complete the following table (1):

| Medium | Type of hemolysis | Other features | Example of grown bacteria |
|-----------------------------|-----------------------|---|---------------------------|
| Inoculated blood agar plate | Plate (A) β | Catalase +ve | S. aureus |
| | Plate (B) α | Catalase -ve aren't inhibited by opterin | Str. Pyogenes |
| | Plate (C) γ | Catalase -ve | Str. bovis |
| | | Plate (D) | |

- Complete the following table (2):

| Medium | Colour of colonies (or other features) | Explanation | Example of grown bacteria |
|--------------------|--|--------------|---------------------------|
| Inoculated N. agar | Plate (A) golden yellow | endo pigment | S. aureus |
| | Plate (B) greenish | exo pigment | Pseudomonas |
| | Plate (C) opaque | | |
| | Plate (D) swarming | motility | Proteus |

- Complete the following table (3):

| Medium | Colour of colonies | Explanation | Example of grown bacteria |
|----------------------|---------------------|---|---------------------------|
| Inoculated MacConkey | Plate (A) Pink | Lactose fermentation | E. Coli ^m |
| | Plate (B) Yellow | Non-lactose fermentation | Salmonella shigella |
| Inoculated MSA | Plate (C) Pink | Non-ferment of mannite | S. epidermidis |
| | Plate (D) Yellow | fermentation of mannite & acid production | S. aureus |
| Inoculated TCBS | Plate (E) Yellow | Sucrose fermentation | 1, 1392 types of cholera |
| | Plate (F) Green | Non-Sucrose fermentation | Other species of cholera |

Report(8): Demonstration of some selected biochemical reactions

ILOs: 1. To examine and identify some selected biochemical reactions.
2. To distinguish positive and negative tests.

1. Catalase test:

- Principle: *differentiation between Staphylococci from Streptococci*

| | Description of | Examples (organisms) |
|----------|---------------------|----------------------|
| +ve test | <i>Has bubbling</i> | <i>Staphylococci</i> |
| -ve test | <i>no bubbling</i> | <i>Streptococci</i> |

2. Oxidase test:

- Principle: *Some bacteria eg Neisseria produce oxidase e. which reduces the oxidase reagent (tetramethyl-P-Phenylene-diamine hydrochloride) to deep purple colour.*

| | Description of | Examples (organisms) |
|----------|---------------------------|----------------------|
| +ve test | <i>deep Purple Colour</i> | <i>Neisseria</i> |
| -ve test | <i>Yellow</i> | <i>Staphylococci</i> |

3. Indole test:

- Principle: Demonstrate ability of bacterium to decompose A.A tryptophan in peptone to produce indole then we test for indole by Kovacs reagent

| | Description of | Examples (organisms) |
|----------|----------------|----------------------|
| +ve test | Red ring | E. coli |
| -ve test | Yellow ring | Klebsiella |

4. Simmonds' citrate test:

- Principle: Demonstrate ability of certain bacteria to utilize citrate as only source of Carbon

| | Description of | Examples (organisms) |
|----------|----------------|----------------------|
| +ve test | blue colour | Klebsiella |
| -ve test | green colour | E. coli |

5. Urease test:

- Principle: Detect Production of urease enzyme in media contain Urea & Phenol red - urease decompose urea & release ammonia so pH alkaline → Phenol red Change to red

| | Description of | Examples (organisms) |
|----------|----------------|----------------------|
| +ve test | deep pink | Proteus - Klebsiella |
| -ve test | Yellow | E. coli - Salmonella |

6. Coagulase test:

- Principle: detects Production of Coagulase enzyme leads to Clotting of Plasma its produced by S. aureus

| | Description of | Examples (organisms) |
|----------|----------------|----------------------|
| +ve test | Clot | S. aureus |
| -ve test | not Clot | any |

7. Methyl red (MR) test:

- Principle: Detect Production of acid in glucose phosphate Peptone

| | Description of | Examples (organisms) |
|----------|----------------|----------------------|
| +ve test | Pink Colour | E. coli |
| -ve test | Yellow Colour | Klebsiella |

8. Voges-Proskauer (VP) test:





- Principle: Detect Production of acetyl methyl Carbinol & small acid in glucose phosphate Peptone during glucose fermentation

| | Description of | Examples (organisms) |
|----------|----------------|----------------------|
| +ve test | Pink Colour | Klebsiella |
| -ve test | Yellow Colour | E. coli |

Triple Sugar Iron agar:

- Composition:
 - Sugars (%): 0.1% glucose 1% lactose 1% sucrose
 - Indicator: Phenol red
 - Ferrous sulphate to detect: for detection H_2S
 - agar agar: - Solidification
- Soft agar cracks on gas Production
- The TSI agar is poured in test tubes in the form of slants with a deep butt
- The medium is of low concentration of agar (soft agar), why?
Cracks on gas Production

- Interpretation of TSI test:
- Complete the following table:

| Drawing of TSI pattern | Butt colour | Slant colour | H_2S and gas | Explanation | Example (organisms) |
|---|-------------|--------------|----------------|--|---------------------|
|  | Red | Red | No | No Carbohydrate fermenting | Pseudomonas |
|  | Yellow | Red | No | ferment glucose only will release small amount of acid | Shigella |
|  | Yellow | Yellow | No | ferment lactose or sucrose release big amount of acid | E. coli |
|  | black | red | ✓ | fermenting glucose only with H_2S Production | Salmonella |

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Objectives of serological diagnostic tests

1. To identify some selected serological tests.
2. To distinguish positive and negative tests and read the titres of positive tests.

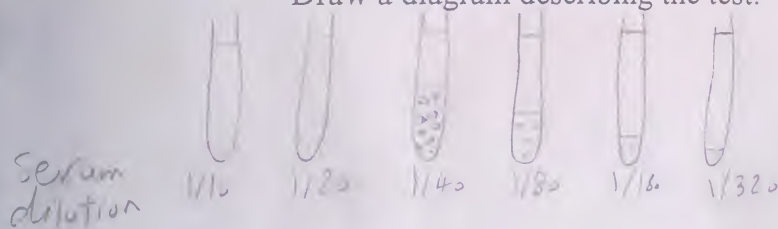
• Tube agglutination test:

1. Widal test:

- Mention antigen suspensions used in this test.
 - *Salmonella* O antigen for *S. dysenteriae*
 - *H* antigen of *S. typhi* *S. Paratyphi A* *S. Paratyphi B*
- Examine the visible clumping at the bottom of the tubes.
- Identify the highest dilution that shows visible agglutination (the titre): ... 1/320.
- Interpretation of the test:
 +ve

2. Brucella standard agglutination test:

- Examine the test and determine the titre: 1/180
- A wide range of dilution of the patients' serum are used. Why?
 to avoid Zone Phenomenon
 if - antibody excess in first
 - IgA blocking antibodies
- Draw a diagram describing the test:



Excess antibody
No Visible reaction

Optimal
 Proportion
 Visible reaction

Excess antigen
 No visible reaction

3. Antistreptolysin O titre:

A test for determination of antibodies titre to streptolysin O of *Strept. pyogenes*

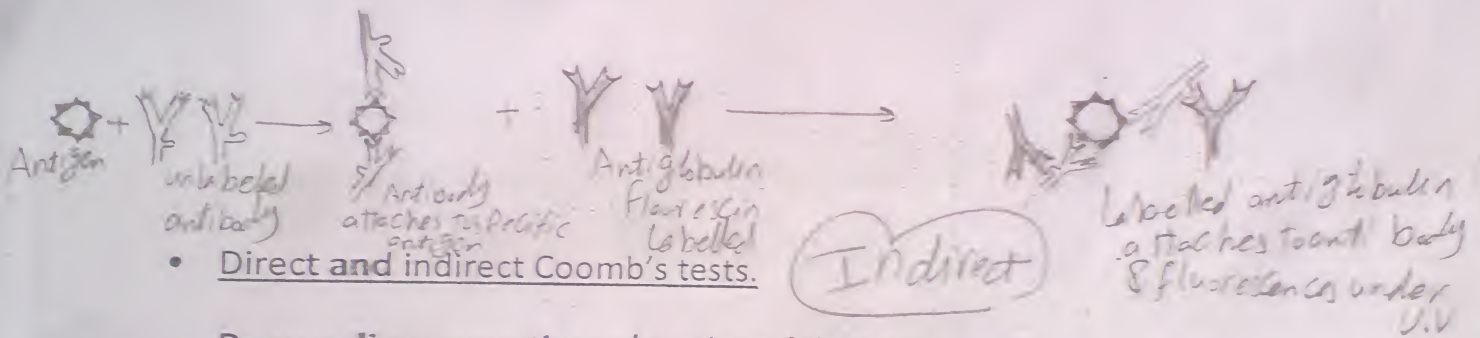
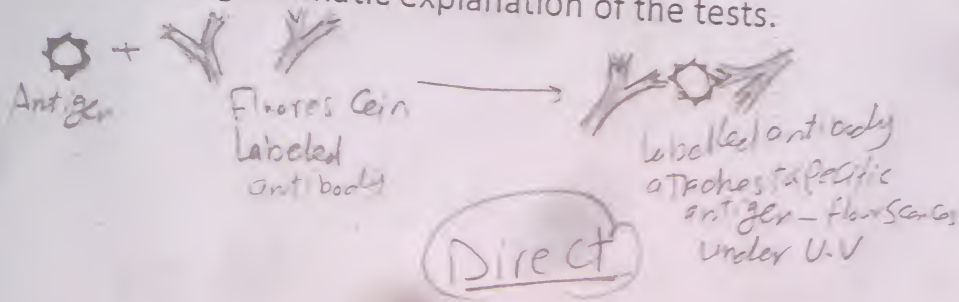
- The highest dilution of the patients' serum showing the ASO titre.
- When should the test is considered positive?
titre more than 1/200
- What is the type of antigen-antibody reaction?
 - Agglutination
 - Precipitation
 - Complement fixation
 - ~~immunofluorescence~~
 - ~~ELISA~~ Toxin anti toxin neutralization

tests based on antigen - antibody reactions and some molecular techniques.

ILOs: To know and understands the principle of the tests.

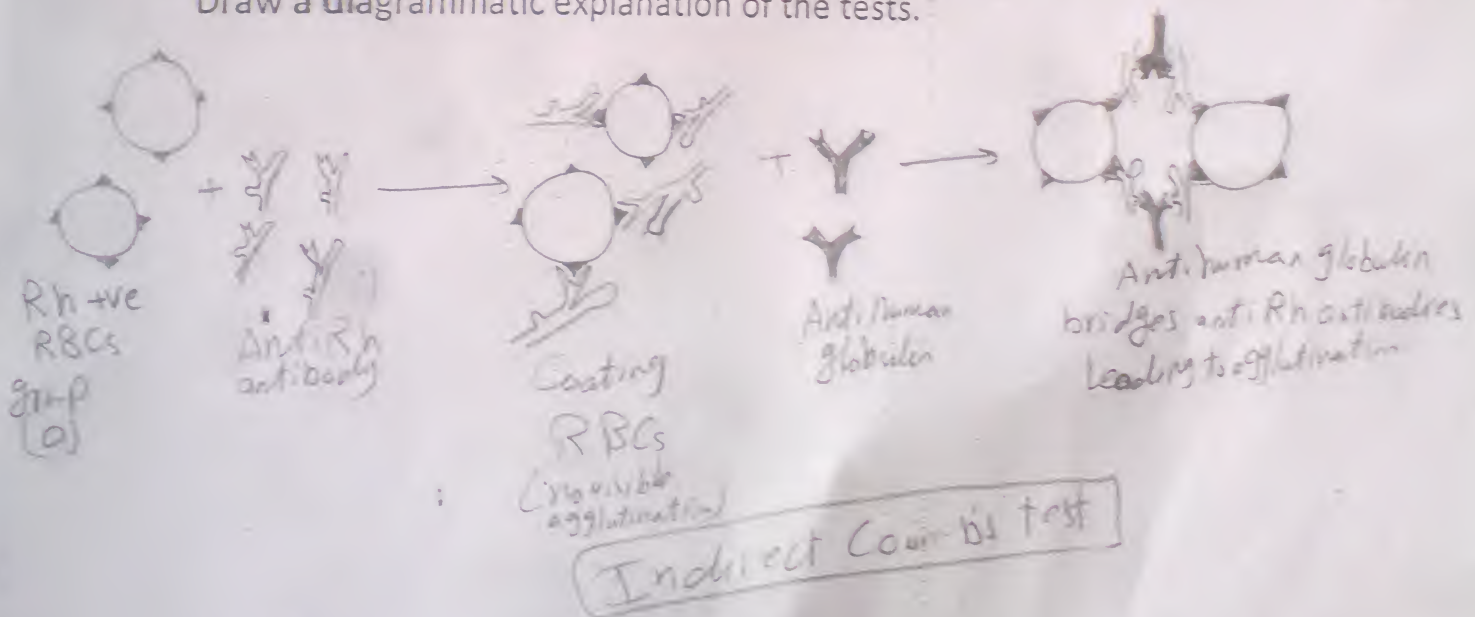
- Direct and indirect florescent technique.

Draw a **diagrammatic** explanation of the tests.



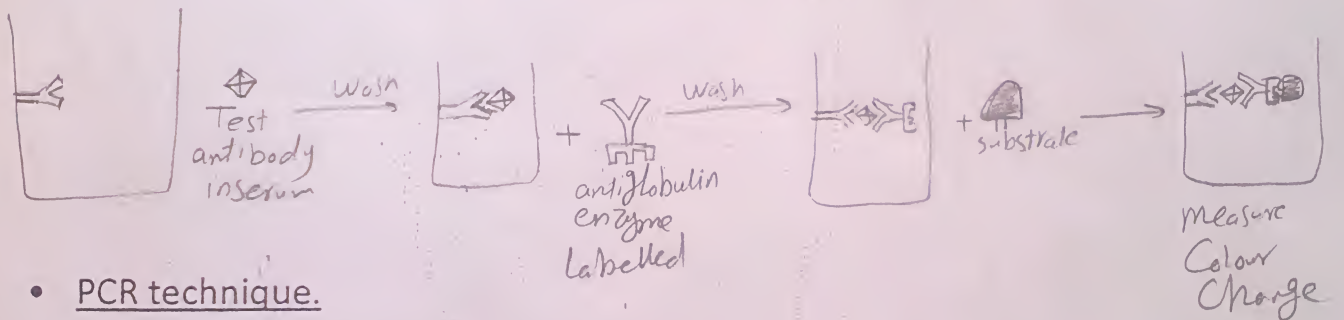
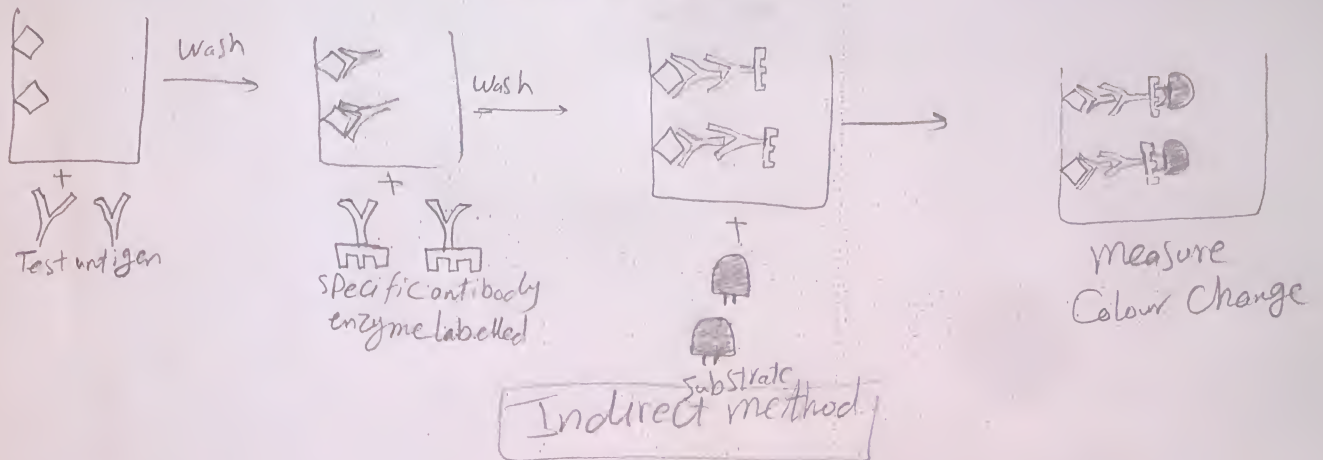
- Direct and indirect Coomb's tests.

Draw a **diagrammatic** explanation of the tests.



- ELISA test.

Draw a diagrammatic explanation of the test.



- PCR technique.

Draw a diagrammatic explanation of the test.

